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TRANSMITTAL OF CERTIFIED PRIORITY DOCUMENT UNDER 35 USC §119

In accordance with the provisions of 35 U.S.C. §119, applicants hereby claim priority of Swedish Patent application No. 0100625-3, filed February 23, 2001. A certified copy of the application is submitted herewith. As the priority application is in the English language, all of the requirements of §119 have been met.

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Respectfully submitted,

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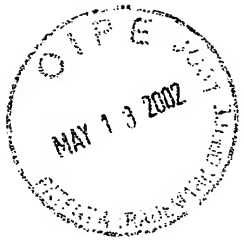

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METHODS FOR PROTEIN PURIFICATION

TECHNICAL FIELD

The present invention relates to a recombinant construct comprising a nucleotide sequence encoding a fusion protein comprising a soluble form of human SSAO (Semicarbazide-Sensitive Amine Oxidase), a secretable fusion partner, a signal peptide; and a protease cleavage site. The invention also relates to methods for purification of a soluble form of human SSAO, said methods utilizing the recombinant construct.

BACKGROUND ART

Semicarbazide-Sensitive Amine Oxidases (SSAOs) belong to the copper-containing amine oxidase family of enzymes (CuAO; EC.1.4.3.6) and are widely distributed among both eukaryotic and prokaryotic organisms (Buffoni, 1993). The physiological role of this abundant enzyme is essentially unknown and endogeneous substrates with high affinity have so far not been identified, although benzylamine is an artificial high-affinity substrate (Buffoni, 1993; Callingham et al., 1995; Lyles, 1996, Hartmann and McIntire, 1997; Holt et al., 1998). In humans high SSAO activity is found in vascular smooth muscle cells (Lewinsohn 1984; Nakos and Gossrau, 1994; Yu et al., 1994; Lyles and Pino, 1998; Jaakkola et al., 1999). SSAO activity has also been detected in smooth muscle cells of non-vascular type and in endothelial cells (Lewinsohn, 1984; Castillo et al., 1998; Jaakkola et al., 1999). Small amounts of SSAO protein is also found in blood showing similar properties compared to the tissue-bound form (Yu and Zuo, 1993; Yu et al., 1994; Kurkijärvi et al., 1998).

Many studies have demonstrated that SSAO activity in blood plasma is elevated in several human conditions such as heart failure, atherosclerosis and diabetes (Lewinsohn, 1984; Boomsma et al., 1997; Ekblom, 1998; Boomsma et al., 1999; Meszaros et al., 1999). The mechanism(s) underlying these alterations of enzyme activity are currently uncharacterized. It has been suggested that reactive aldehydes and hydrogen peroxide produced by endogenous amine oxidases could be causative or

contribute to the progression of cardiovascular diseases, and that inhibition of SSAO activity in diabetics might decrease vascular complications (Ekblom, 1998).

Recently it was found that the cDNA sequence of human SSAO (Zhang and McIntire, 1996) is identical to the vascular adhesion protein 1 (VAP-1), which participates in lymphocyte recirculation by mediating the binding of lymphocytes to peripheral lymph node vascular endothelial cells (Smith et al., 1998; see also WO 98/53049). The cDNA sequence of SSAO / VAP-1 is deposited under GenBank Accession Nos. U39447 and NM_003734 (SEQ ID NO: 1). VAP-1 has also been found to be up-regulated on the endothelial cell surface under inflammatory conditions (Smith et al., 1998). However, the adhesive properties of SSAO have only been found in endothelial cells. In smooth muscle cells, SSAO does not support binding of lymphocytes (Jaakola et al., 1999). DNA-sequence analysis, structural modeling and experimental data suggest that human SSAO is a homodimeric glycoprotein consisting of two 90-100 kDa subunits anchored to the plasma membrane by a single N-terminal membrane spanning domain (Morris et al., 1997; Smith et al., 1998; Salminen et al., 1998).

No reports have so far been published regarding the purification of a recombinant mammalian SSAO or purification to near homogeneity of larger amounts of a human SSAO from a natural source. One report has described the use of a FLAG peptide fused to the N-terminal end of full-length human SSAO for detection purposes, but no results were presented regarding its use for purification of the human SSAO protein (Smith et al., 1998). Monoclonal antibodies have been used to immunoaffinity purify small amounts of human SSAO from serum and tissue homogenates for immunoblotting (Smith et al., 1998; Kurkijärvi et al., 1998). Consequently, there is a need for alternative methods for the purification of human SSAO in significant amounts.

Glutathione S-transferase (GST) from *Schistosoma japonicum* is a homodimeric cytoplasmic enzyme that can be purified by affinity chromatography using immobilized cofactor glutathione, followed by competitive elution using reduced glutathione (GSH). Taking advantage of this specific interaction, a gene fusion system for *E. coli* intracellular expression was developed by Smith and co-workers (Smith & Johnson, 1988; see also WO 88/09372) to facilitate detection and purification of recombinant

proteins fused to GST. A potential drawback with using GST as fusion partner is the possibility that the free cysteines on its surface can crosslink with free cysteines on e.g. the fused target protein when exposed to an oxidizing environment. To minimize this risk and to allow for secretion of GST-fusion proteins a mutant form of GST was recently developed, which retain both its ability to form homo-dimers and its enzyme activity (Tudyka and Skerra, 1997). The homo-dimerization propensity of GST can be used to provoke dimerization of the fused target protein e.g. for the purpose of increased avidity effects (Tudyka and Skerra, 1997).

Alternative homodimeric fusion partners described in the literature are e.g. the Fc region of immunoglobulins (Hollenbaugh et al., 1992; Sakurai et al., 1998; Lo et al., 1998; Dwyer et al., 1999) and leucine zippers such as GCN4 (Rieker and Hu, 2000; Müller et al., 2000). Several different proteins have been fused to these homodimeric protein domains for different purposes e.g. to increase avidity (Dwyer et al., 1999; Muller et al., 2000) and to restore high-affinity DNA binding of truncated DNA-binding proteins (Rieker and Hu, 2000). Fc-fusion protein can be purified by protein A-affinity chromatography involving elution with low pH buffers (Sakurai et al., 1998; Lo et al., 1998), which may decrease activity of the fused target protein (Gräslund et al., 1997). Another problem associated with using Fc as fusion partner is the use of serum for cell growth, which complicate detection and purification of secreted Fc-fusions since serum contains large amounts of immunoglobulins (Sakurai et al., 1998). The leucine zipper GCN4 has mostly been used as fusion partner for proteins expressed in *E. coli* (Müller et al., 2000) and an affinity-tag might has to be fused to facilitate purification.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1

Schematic illustration of the GST-SSAO DNA construct. The three cysteine to serine mutations (residues 85, 138, and 178 according to the sequence having GenBank Accession No. M14654) in the GST fusion partner are shown with boldface letters. Boxed sequence represents the recognition sequence for the 3C-protease.

Fig. 2

Overview of the SSAO purification process. The determined specific activities for each purification step are indicated.

Fig. 3

Schematic illustration of the GST-SSAO expression vector designated pMB887.

DISCLOSURE OF THE INVENTION

According to the present invention, it has unexpectedly been found that soluble human SSAO can be produced in milligram quantities in a purification system utilizing a fusion partner capable of enabling dimerization of soluble SSAO. Consequently, in a first aspect this invention provides a recombinant construct comprising a nucleotide sequence encoding a fusion protein comprising

(i) a soluble form of human SSAO;

(ii) a secretable fusion partner enabling dimerization of SSAO;

(iii) a signal peptide allowing for secretion of a polypeptide from a host cell into the culture medium; and

(iv) a protease cleavage site located between the human SSAO variant and the fusion partner.

As will be understood by the skilled person, the recombinant construct can optionally comprise one or more nucleotide sequences coding for spacer amino acid sequences of various lengths. Such spacer sequences could be used in order to increase the flexibility within the fusion protein, or to increase the space between protein domains so that folding can take place independently of adjacent domains. Further, spacers could be useful for increasing the accessibility for a protease to cleave at an introduced cleavage recognition sequence.

The said soluble form of human SSAO is preferably lacking the membrane spanning portion of wild-type human SSAO. The membrane spanning portion of the SSAO

polypeptide is known in the art (Morris et al., 1997; Holt et al., 1998; Smith et al., 1998) and is essentially set forth as amino acids 5 to 27, in particular amino acids 6 to 26, of SEQ ID NO: 2.

The amino acid sequence for human SSAO, excluding the membrane spanning portion, preferably comprises, or essentially consists of, positions 29 to 763 in SEQ ID NO: 2. However, the skilled person will understand that a part of the membrane-spanning portion could be included in the SSAO polypeptide while the polypeptide would still retain its essentially soluble properties. Consequently, the amino acid sequence for human SSAO could comprise e.g. positions 27 to 763, or 28 to 763, of SEQ ID NO: 2, including fragments thereof having substantially the biological activities of human SSAO. Further, the term "human SSAO polypeptide" is intended to encompass mutants and naturally occurring variants of human SSAO, either having retained enzymatic activity or protein interaction (e.g. adhesion function), or designed to facilitate structural studies (e.g. improved properties for crystallization), or mutated to facilitate studies of structure/function relationships (which also includes inactive mutants).

The fusion partner can be fused to the C-terminal or N-terminal portion of the human SSAO polypeptide. It is envisaged that the fusion protein could comprise more than one fusion partner, for instance one fused to the N-terminal and one fused to the C-terminal part of SSAO. An additional fusion partner could be an additional affinity tag, or a reporter protein such as Enhanced Green Fluorescent Protein (EGFP).

A large number of different gene fusion systems and fusion partners have been described. In such systems, different types of interactions, such as enzyme–substrate, bacterial receptor–serum protein, polyhistidines–metal ion, and antibody–antigen, have been utilized (Uhlén et al., 1992). Various gene fusion systems for affinity purification are also known in the art. Examples of fusion partners used in such systems (for reviews, see e.g. Nilsson et al., 1997; or Sheibani, 1999) comprise staphylococcal Protein A and its derivative Z; the albumin-binding protein from streptococcal Protein G; glutathione S-transferase (GST); polyhistidine tags; biotinylated affinity tags (e.g Biotin AviTag); *E. coli* maltose-binding protein; cellulose binding domains; the FLAG peptide; and *Strep*-tag. Alternative systems may be engineered using protein scaffolds

for generation of novel ligand receptors (see Skerra, 2000, and references therein). These novel binding proteins, e.g. affibodies, may then be useful as fusion partners for different applications (Nygren and Uhlén, 1997; Nord et al., 1997).

According to this invention, the said fusion partner should enable dimerization of SSAO. A suitable fusion partner is glutathione S-transferase (GST), because of its propensity to dimerize and because the purification procedure has the potential to be performed under mild conditions using chromatography media with immobilized glutathione (e.g. from Amersham Pharmacia Biotech, Uppsala, Sweden). In addition, GST can conveniently be detected either by its enzymatic activity or by the use of GST specific antibodies or glutathione, using commercially available GST detection systems (e.g. from Amersham Pharmacia Biotech). The fusion partner could also be a functionally equivalent variant of GST, having retained propensity for dimerization and having binding properties allowing affinity purification. The said fusion partner is more preferably a variant of *S. japonicum* GST (GenBank Accession No. M14654; SEQ ID NOS: 3 and 4), designed for secretion out of the host cell, having one or more of the cysteine residues in positions 85, 138, and 178 replaced with other amino acid residue(s). Most preferably, the said variant has all the cysteine residues in positions 85, 138, and 178 replaced with serine residues (see Tudyka & Skerra, 1997 and SEQ ID NO: 5).

In addition, the said recombinant construct should comprise a nucleotide sequence encoding an N-terminal signal peptide, which allows for secretion of the said fusion protein from a host cell into the culture medium. For production of a human protein such as SSAO in a eukaryotic cell a homologous signal peptide is preferred. For production of SSAO in HEK293 cells e.g. a mouse IgG1 heavy chain signal peptide (Kabat et al., 1991) may be used. Other suitable signal peptides are known in the art and are described in e.g. Kabat et al., *supra*.

Several methods have been described for site-specific cleavage of fusion proteins based on treatment with chemical agents such as CNBr or hydroxylamine, or enzymes such as enterokinases, Factor Xa, thrombin, subtilisin or other proteases (see e.g. Nilsson et al. (1997) and references therein). According to this invention, the said fusion partner can

conveniently be removed from human SSAO by protease cleavage. The protease to be used for cleavage can e.g. be a 3C protease from the picornavirus family, e.g. a rhinovirus or enterovirus 3C protease (Walker et al., 1994). Consequently, the protease cleavage site can preferably be a cleavage site for a 3C-protease from the picornavirus family, e.g. a rhinovirus or enterovirus 3C protease. In one exemplified form of the invention, the said 3C protease cleavage site comprises the amino acid sequence EALFQG. However, the skilled person will be able to identify other suitable cleavage sites, see e.g. Blom et al. (1996) and references therein.

The recombinant construct according to the invention could e.g. comprise a nucleotide sequence encoding essentially the amino acid sequence shown in Figure 1. The invention also provides an expression vector, prepared according to standard methods, comprising the recombinant construct according to the invention. Such an expression vector is exemplified by the expression vector termed pMB887, shown in Figure 3.

In another aspect, the invention provides a method for the purification of a recombinant human SSAO polypeptide comprising the steps of:

(i) transfecting cells with an expression vector according to the invention, as defined above;

(ii) culturing the said cells under conditions allowing for the fusion protein expressed by the vector to be secreted into the cell medium;

(iii) binding the obtained fusion protein to a medium comprising a ligand having affinity for the fusion partner;

(iv) separating the said fusion partner and the SSAO polypeptide; and

(v) recovering the purified human SSAO polypeptide.

The fusion partner can be separated from the human SSAO variant either when the fusion protein is still attached to the affinity ligand, or when the fusion protein has been released from the affinity ligand. When the said fusion partner is GST, the said ligand having affinity for the fusion partner is preferably glutathione, or a derivative thereof. Alternatively, antibodies directed to GST could be used as affinity ligands.

As mentioned above, the fusion partner can be separated from human SSAO by protease cleavage with e.g. a picornavirus, such as rhinovirus, 3C-protease. The said protease can be fused to a fusion partner, thereby obtaining a "fusion protease" (see Walker et al., 1994; Gräslund et al., 1997). Such a fusion partner can conveniently be the same fusion partner as used for the SSAO polypeptide, e.g. glutathione S-transferase. However, other suitable fusion partners for proteases, such as albumin-binding protein from streptococcal Protein G, are known in the art, see e.g. Gräslund et al., 1997. The said fusion protease can be separated from the SSAO polypeptide by a process comprising binding the fusion protease to a medium comprising a ligand having affinity for the said fusion partner. Consequently, when the fusion partner is GST, the said ligand is preferably glutathione, or a derivative thereof. As mentioned above, antibodies directed to the fusion partner could also be used as affinity ligands. A commercially available system is the PreScission Protease (Amersham Pharmacia Biotech,) which is a genetically engineered fusion protein consisting of *S. japonicum* GST and human rhinovirus 3C protease.

For certain application, it might be advantageous to have SSAO immobilized. This may be achieved e.g. by an affinity-tag such as GST as described above. Examples of applications where a fusion protein is immobilized via an affinity-tag include: capture of protein ligands, analysis of protein-protein interactions, and use in bioreactors (Nilsson et al., 1996; Nord et al., 1997; Shpigel et al., 1999). However, many alternative methods for protein immobilization are described (see e.g. Tischer and Kasche, 1999, and references therein), that also may be applicable for immobilization of GST-SSAO or SSAO after removal of the fusion partner, such as covalent binding and non-covalent adsorption. In addition, the SSAO protein might also be encapsulated in e.g. sol-gel or an artificial cell e.g. a liposome (see e.g. Liang et al., 2000, and references therein). One advantage with an affinity-tag such as GST is that an oriented immobilization can be achieved, often in a one-step procedure directly from e.g. a cell lysate (Nilsson et al., 1997; Saleemuddin, 1999). This may result in good steric accessibility of active binding sites and increased stability (Saleemuddin, 1999; Turkova, 1999). Examples of alternative affinity-tag approaches that has been used for immobilization of proteins are e.g. peptides and proteins that can be specifically biotinylated by biotin ligase and used as fusion partners to take advantage of the very strong interaction ($K_d \sim 10^{-15}$) between

biotin and streptavidin or avidin (Nilsson et al., 1997), and CBDs which binds specifically to cellulose (Linder et al., 1998; Tomme et al., 1998). Oriented immobilization of a protein may also be achieved by using immobilized antibodies that binds the protein or through carbohydrate moieties that may be present on the protein surface (Turkova, 1999).

Recently, amine oxidase from pea seedlings was immobilized using a modified carbon paste to yield a biosensor for determination of biogenic and synthetic amines (Wimmerova and Macholan, 1999). Similarly, recombinant human SSAO might be immobilized for construction of biosensors to detect e.g. the cardiovascular toxin allylamine which is used in industrial organic processes and is a substrate for SSAO (Boor and Hysmith, 1987; Conklin et al., 1998). When immobilized, recombinant SSAO may be envisioned to mimic a membrane-anchored SSAO and its characteristics, which might differ from the soluble state.

Consequently, as shown in the following examples, the invention provides a procedure for the production of a highly purified soluble recombinant human SSAO with enzymatic activity. The exemplified procedure involves the use of a mutant form of *S. japonicum* glutathione *S*-transferase (GST), designed for transport out of the host cells (Tudyka and Skerra, 1997), as an affinity fusion partner. The fusion protein was secreted from mammalian cells and could be purified directly from the culture medium by glutathione-affinity chromatography. By specific proteolysis and an additional glutathione-affinity chromatography step, the fusion partner and the protease were removed, whereby pure, soluble and highly active recombinant human SSAO protein was obtained in milligram quantities. To the inventors' knowledge, this is the first time an active recombinant soluble form of the human SSAO protein has been produced and purified to near homogeneity.

It is believed that the disclosed process for production of recombinant human SSAO will be applicable also to other mammalian amine oxidases, such as the human placenta diamine oxidase (Zhang et al., 1995) and the human retina-specific amine oxidase (Imamura et al., 1998), as well as for other secretable proteins. The disclosed process may also facilitate the discovery and identification of modifications e.g. the

identification of the active site cofactor, e.g. by isolation of cofactor-containing peptides or by crystal structure determination.

In the following examples, it is shown that SSAO is active and soluble without its transmembrane region, and that GST can be proteolytically removed. These findings support the hypothesis that SSAO is released into circulation by proteolytic cleavage near the transmembrane region (shedding), a process which is common for Type I and Type II membrane proteins (Hooper et al., 1997). The elevated SSAO activity in plasma in e.g. diabetes (Boomsma et al., 1999) may thus be the consequence of increased proteolytic activity of a protease that cleave the membrane-anchored SSAO, or of increased surface localization increasing the substrate availability for an existing protease.

EXPERIMENTAL METHODS

PCR-amplification and cloning of the human SSAO gene from aorta cDNA

Two PCR-primers were designed with the help of the published cDNA sequence of human placenta amine oxidase (GenBank Accession No. U39447; Zhang and McIntire, 1996). The 5'-primer XNQZ-15 (5'-CCG GAA TTC CAA CGC GTC CAT GAA CCA GAA GAC AAT CCT CGT G-3') was designed to hybridize to the 5'- end of the SSAO coding sequence including the ATG start codon and to contain the restriction enzyme cleavage sites *Eco*RI and *Mlu*I for cloning. The 3'-primer XNQZ-17 (5'-CCC CCA AGC TTG TCG ACT CAC TAG TTG TGA GAG AGA AGC CCC CCC-3') was designed to hybridize to the 3'-end including the native stop codon TAG followed by an additional stop codon TGA and two restriction enzyme cleavage sites for cloning, *Sal*I and *Hin*dIII. As template for the PCR 0.5 µl human aorta or human smooth muscle cell QUICK-Clone cDNAs (1ng/µl, Clontech Laboratories, Palo Alto, CA) were tested. The following conditions was used for the PCR-reaction, 20 pmol of each primer XNQX-15 and XNQZ-17, 1 µl dNTPs (10 mM), 1 µl Advantage cDNA Polymerase Mix (Clontech), 5 µl 10x cDNA PCR reaction buffer (Clontech) in a total volume of 50 µl.

Amplification was performed with a Perkin-Elmer 2400 thermocycler (Perkin-Elmer, Norwalk, CT). The PCR-program consisted of an initial denaturation at 94°C for 5 min, 35 cycles of 94°C for 30 s, 60°C for 30 s and 72°C for 3 min followed by a final extension at 72°C for 3 min. TA-cloning was then used to insert the PCR-product into the vector pCR2.1-TOPO (Invitrogen, Carlsbad, CA). The cloned PCR-fragment was sequenced in both directions according to a standard protocol for dye terminator cycle sequencing and analyzed on a DNA sequencer ABI 377 (Applied Biosystems, Foster City, CA).

Construction of vectors for expression of SSAO in mammalian cells

A vector for expression of the complete SSAO protein in mammalian cells was prepared by insertion of the *Eco*RI and *Sal*I fragment from the pCR2.1TOPO-SSAO vector into the same sites of the vector pCI-neo (Promega, Madison, WI), resulting in the vector pMB843. This vector was used as template for PCR-amplification of the region corresponding to residues 29-763 of the human SSAO (Zhang and McIntire, 1996). A 5'-primer 5'- GAG GAA GCT TTG TTC CAA GGT GGA GAT GGG GGT GAA-3' was synthesized containing codons for a partial 3C protease cleavage site (see below) and a *Hin*dIII restriction enzyme cleavage site upstream of the codon for residue 29. The 3'-primer 5'-GCA TTC TAG TTG TGG TTT GTC-3' is a pCI-neo vector specific primer annealing downstream of the cloned SSAO fragment. The resulting PCR-product was digested with *Hin*dIII and *Not*I and cloned into the plasmid pET38b(+) (Novagen, Inc., Madison, WI) cut with same enzymes, resulting in pET38-SSAO. DNA sequencing was performed as described above to verify expected sequence of the cloned SSAO fragment.

A mutated form (SEQ ID NO: 5) of the glutathione S-transferase (GST) from *S. japonicum* previously used as a secretable enzymatically active dimerization module for a recombinant protein (Tudyka and Skerra, 1997) was prepared by PCR-mediated mutagenesis and assembly of fragments as described below. The mutations was performed to replace three cysteine residues 85, 138, and 178 located close to the GST protein surface as revealed in the crystal structure of the *S. japonicum* GST (Lim et al., 1994; Tudyka and Skerra, 1997) with serine residues in order to avoid unwanted

disulphide formation after export of the GST fusion protein to an oxidizing environment (Tudyka and Skerra, 1997). The following PCR-primers were used to construct the mutated GST and to introduce the first part of a 3C protease cleavage site (see below) as well as suitable restriction enzyme cleavage sites for cloning. In addition, the primers introduce internal restriction sites for control cleavage and for possibility to assemble PCR-fragments by ligation: ROEL-1 (5'-GCC GGA ATT CGA CGC GTC CCC TAT ACT AGG TTA TTG G-3') contains *Eco*RI and *Mlu*I for cloning and anneals to codons 2-8 of GST (M14654); ROEL-2 (5'-CTC TGC GCG CTC TTT TGG AGA ACC CAA CAT GTT GTG C-3') contains a *Bss*HII site; ROEL-3 (5'-GGT TCT CCA AAA GAG CGC GCA GAG ATT TCA ATG CTT GAA G-3') contains a *Bss*HII site; ROEL-4 (5'-ATG AGA TAA ACG GTC TTC GAA CAT TTT CAG CAT TTC-3') contains a *Bbs*I site; ROEL-5 (5'-GTT CGA AGA CCG TTT ATC TCA TAA AAC ATA TTT AAA TGG TGA TC-3') contains a *Bbs*I site; ROEL-6 (5'-AAA AGA AAC TAG TTT TGG GAA CGC ATC CAG GCA-3') contains a *Spe*I site; ROEL-7 (5'-CCC AAA ACT AGT TTC TTT TAA AAA ACG TAT TGA AGC TAT C-3') contains a *Spe*I site; ROEL-8 (5'-ACC CAA GCT TCC TGA CTT TGT GAC TTT GGA GGA TGG TCG CCA CC-3') contains *Hin*dIII for cloning and anneals to codons 212-218 of GST (M14654). ROEL-8 will also introduce codons for a spacer-sequence SQSQ before a partial 3C protease cleavage site. Overlapping parts of the GST gene were amplified in separate PCR-reactions with primer pairs ROEL-1/2, ROEL-3/4, ROEL-4/5 and ROEL-7/8, using plasmid pGEX-6P-2 (Amersham Pharmacia Biotech) as template. This allowed the complete mutated GST gene to be assembled by mixing the four PCR-fragments and using them as templates in a PCR reaction with primers ROEL-1 and ROEL-8. The PCR-reactions was performed using the Advantage cDNA PCR Kit (Clontech). In the next step the GST fragment was digested with *Eco*RI and *Hin*dIII and cloned into the same sites of pUC18 (Amersham Pharmacia Biotech), yielding pMB809. DNA sequencing was performed as described above to confirm the expected sequence of the mutated GST fragment. The pMB809 vector was cleaved with *Eco*RI and *Hin*dIII and the GST fragment was isolated and cloned upstream of the SSAO fragment in the pET38-SSAO vector cut with the same enzymes. This step resulted in the creation of a complete 3C protease cleavage site EALFQG of human rhinovirus-14 and coxsackievirus (Miyashita et al., 1996; Wang et al., 1997) between GST and SSAO (residues 29-763) (see Fig. 1).

The GST-SSAO fragment was cloned in the *Mlu*I and *Sal*I site of the vector pMB565, in which a mutated signal sequence of a murine IgG1 heavy chain (Fig. 1) is cloned in the multilinker of the mammalian expression vector pCI-neo (Promega). The resulting GST-SSAO expression vector was named pMB887 (Fig. 3).

Transfection and selection of stable clones

Three 25 cm^2 T-flasks were seeded with approximately $4x10^5$ human embryo kidney 293 cells (HEK293 cells, ATCC CRL–1573, Rockville, MD). Cells were grown to ~50% confluency in growth medium containing Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 2 mM L-Glutamine. The FBS was heat-inactivated at 56°C for 30 min before mixed with the growth medium components. The expression vector pMB887 was then introduced into the cells by liposome-mediated transfection using LipofectAMINE according to the manufacturer's recommendations (Life Technologies, Frederick, MD). After 48 hours of growth the medium was changed in all flasks to growth medium supplemented with 1 mg/ml geneticin (G418) for selection of stably transfected cells. Approximately two weeks later, resistant cells emerged and were grown to confluency. Cells from the three T-flasks were pooled (clone-mixture) and diluted in growth medium supplemented with 1.2 mg/ml G418 and seeded in a 15-cm Petri dish. Individual colonies emerged after two weeks and were picked to be expanded individually for subsequent analysis of GST-SSAO production. Detection of GST proteins in collected medium from expanded clones was performed using the GST 96-Well Detection Module (Amersham Pharmacia Biotech). Seven positive clones were selected and frozen.

Production of GST-SSAO in Cell Factories

Clone number 10 was expanded and cultured in growth medium containing DMEM supplemented with 5 % FBS (heat-inactivated), 2 mM L-Glutamine and 1.2 mg/ml G418 and used to seed a 6320 cm^2 Nunc Cell Factory (Nalge Nunc Int., Naperville, IL) containing 1500 ml of growth medium and grown at 37°C. After four days of growth, cells were confluent and medium was collected. New growth medium (1500 ml) with

reduced amount of FBS (2%) was then added to the cells in the same Cell Factory followed by harvest of conditioned medium after three days. This procedure was repeated once resulting in a total of ~4.5 liters of harvested medium from one Cell Factory. Collected medium was centrifuged and stored at –70°C.

Concentration of conditioned cell medium

Frozen conditioned medium from two Cell Factories (9.4 liters) was thawed in a water-bath at 30°C. The material was pumped through an Omega membrane (MWCO (Molecular-Weight Cut-Off) 10000) using a Centramate ultra-filtration equipment (Pall Filtron, Northborough, MA), until a volume of 600 ml was achieved. The retentate was filtered through a 0.45 µm filter, Sartobran P, equipped with a 0.65 µm prefilter (Sartorius, Göttingen, Germany). Remaining filtrate in tubings was displaced by 250 ml of phosphate-buffered saline (PBS) yielding 850 ml of filtered sample.

Purification and cleavage of GST-SSAO

The GST-SSAO fusion protein was purified by glutathione-affinity chromatography on a HR 10/10 column (Amersham Pharmacia Biotech) packed with 8 ml glutathione-Sepharose 4 Fast Flow (Binds ~10 mg GST/ml gel, Amersham Pharmacia Biotech), equilibrated with 10 column volumes of PBS. The filtered material (850 ml), was loaded at 0.9 ml/min over night at room temperature. Flow-through material was collected for analysis and stored at –20°C. After washing the column with PBS, bound proteins were eluted with elution buffer (20 mM GSH, 0.1 M NaCl, 0.1 M Tris-HCl, pH 8.2).

The eluate was loaded on a HiPrep Desalt 26/10 column (Amersham Pharmacia Biotech) equilibrated with helium-sparged cleavage-buffer (150 mM NaCl, 1 mM EDTA, 50 mM Tris-HCl, pH 7.5 at 25°C) and the protein peak was collected. Cleavage was started by adding DTT (dithiothreitol) to 5 mM and 380 units of PreScission protease (Amersham Pharmacia Biotech). The PreScission Protease is a genetically engineered fusion protein consisting of GST and human rhinovirus 3C protease and

cleaves specifically between the Gln (Q) and Gly (G) residues of its recognition sequence.

The cleavage mixture was incubated at 5°C. After 63 hours of incubation the material was loaded on a glutathione-Sepharose column as described above, equilibrated with cleavage buffer. The flow-through (36 ml) was collected and stored at 5°C for approximately one week in an open tube. Samples were withdrawn and analyzed by SDS-PAGE (non-reducing). Proteins captured on the column were eluted with elution buffer for analysis. The collected protein sample was applied on a JumboSep device (MWCO 30000, Pall Filtron) for buffer exchange and concentration. Five cycles of centrifugation and dilution with a buffer containing 50 mM Tris-HCl (pH 7.5) and 150 mM NaCl were performed. A sample was taken for different analyses. The buffer exchanged and concentrated material (4.2 ml) was then stored at –70°C.

Protein analyses

The purification and size of SSAO were analyzed by SDS-PAGE. Samples were electrophoresed in the presence or absence of 2-mercaptoethanol in gradient gels 4-20% or 4-12% (Novex, Copenhagen, Denmark) and proteins were visualized by Coomassie staining (PhastGel Blue R, Amersham Pharmacia Biotech). Protein concentrations were determined with Coomassie Plus protein assay reagent kit (Pierce, Rockford, IL) in 96-well plates with bovine serum albumin as standard according to the manufacturer's procedure.

Size exclusion chromatography was performed on a Superdex 200 PC 3.2/30 column (Amersham Pharmacia Biotech) using the SMART System (Amersham Pharmacia Biotech). The column was equilibrated at room temperature with a buffer containing 50 mM Tris-HCl (pH 7.5), 150 mM NaCl and 1 mM EDTA. Injection volume was 10 µl and samples were eluted at a flow rate of 0.1 ml/min. For column calibration molecular weight markers Blue Dextran 2000 (~2000 kDa), thyroglobulin (669 kDa), ferritin (440 kDa), catalase (232 kDa) and aldolase (158 kDa) from the Gel Filtration HMW Calibration Kit (Amersham Pharmacia Biotech) was used.

N-terminal sequencing was performed on purified GST-SSAO and SSAO by repeated Edman degradation using a HP G1000A protein sequencer coupled to a HP 1090 PTH analyzer (Hewlett Packard, Palo Alto, CA). The GST-SSAO sample was desalted to remove glutathione prior to analysis. SSAO was taken from the flow-through of the glutathione-Sepharose column after cleavage.

A spectrophotometric assay for monoamine oxidases described by Holt and coworkers (Holt et al., 1997) was used to determine amine oxidase activity in samples from the different purification steps. The assay was performed in 96-well microtiter plates incubated at 37°C in a SPECTRAmax 250 microplate spectrophotometer (Molecular Devices, Sunnyvale, CA). The reagent mix containing 1 mM vanillic acid (Sigma, St. Louis, MO), 500 µM 4-aminoantipyrine (Sigma), and 4U ml^{-1} peroxidase (type VI from horseradish, Sigma) in 0.2 M potassium phosphate buffer (pH 7.6) was prepared on the same day assays were performed and kept at 5°C until used. Reactions were started by mixing 50 µl sample, 50 µl reagent mix and 200 µl potassium phosphate buffer with or without 750 µM benzylamine hydrochloride (Sigma) and were performed in triplicate. In order to obtain blank reference values, wells were analyzed with buffer added in place of sample. Absorbance changes were followed at 490 nm for 10-40 minutes. Standard curves were prepared with dilutions of a stock solution of H_2O_2 in potassium phosphate buffer ranging from 10 nmol/well to 120 nmol/well. When inhibition experiments were carried out, the samples were incubated in 300 µM semicarbazide at 37°C for 30 minutes, before addition of the benzylamine solution.

EXAMPLES

EXAMPLE 1: Cloning of SSAO cDNA

A PCR-strategy was used to amplify the gene of a human SSAO from human aorta cDNA. The PCR-primers were designed to include sequences flanking the human placenta amine oxidase gene (Zhang and McIntire, 1996) and to include restriction enzyme cleavage sites for cloning into different expression vectors. The ~2300 bp PCR-product was cloned and subsequent DNA-sequencing showed that the sequence of the

cloned PCR-product was identical to the human placenta amine oxidase sequence (Zhang and McIntire, 1996) and to the VAP-1 sequence cloned from lung cDNA (Smith et al., 1998).

EXAMPLE 2: Purification of membrane-bound SSAO (Example for comparison)

Attempts to produce a recombinant SSAO protein showed that active SSAO could be produced in human embryo kidney (HEK293) cells using the pMB843 vector in which the entire coding sequence of human SSAO was cloned. Active protein was found after extraction using solubilizing agents, but only microgram amounts of protein could be partially purified.

EXAMPLE 3: Rationale and design of a gene construct for expression of a soluble form of SSAO

An alternative strategy was developed for production of a non-membrane-bound SSAO in mammalian cells. Purification and detection were performed by replacing the N-terminal region containing the putative membrane spanning peptide with an affinity fusion partner having an inherent dimerization propensity. The strategy also involved the use of a secretable affinity fusion partner to be able to secrete the fusion protein into the culture medium. A mutated variant of *S. japonicum* glutathione S-transferase (GST) was selected. This mutant GST retains its activity as well as its propensity to dimerize and have been optimized for secretion (Tudyka and Skerra, 1997).

A protease cleavage site was designed to enable release of SSAO from the purified GST-SSAO fusion protein. Scanning of the predicted amino acid sequence revealed an arginine at position 28 flanked by three glycine residues. Several human proteases cleave after basic residues (Carter, 1990; Hooper et al., 1997) and short stretches of glycine residues have been suggested to enhance accessibility to proteases (Carter, 1990). In addition, the proteolytic release of the extracellular region (shedding) of many membrane-anchored proteins into the blood stream occurs close to the membrane

(Hooper et al., 1997). The glycine residue at position 29 was therefore chosen to be linked to a suitable substrate for site-specific proteolysis after purification of the GST-SSAO fusion protein. Thus, a protease that can cleave a substrate having a glycine in the P1' position and having high specificity was desired. Several commercial proteases exist having these two properties such as factor Xa, thrombin, enterokinase and 3C protease (Nilsson et al., 1997). The ability to easily capture the protease after cleavage was another factor considered, leading to the selection of a commercially available 3C protease fused to GST. The 3C protease cleavage site EALFQG (Miyashita et al., 1996; Wang et al., 1997) was introduced in the GST-SSAO fusion construct (Fig. 1).

The GST-SSAO fragment was cloned in frame with a signal sequence to achieve secretion of the GST-SSAO fusion protein into the culture medium. A signal sequence derived from the heavy chain of a murine antibody was used (see Fig. 1). The final construct thus encoded a fusion protein comprising of an antibody signal peptide, an 18 amino acid spacer region, the mutated GST protein, a substrate sequence for the 3C protease and residues 29-763 of the human SSAO protein cloned from human aorta cDNA (Fig. 1). The calculated molecular weight of the unmodified GST-SSAO fusion protein is 112 kDa.

EXAMPLE 4: Initial analyses on conditioned medium from HEK293 cells transfected with the GST-SSAO expression vector

Benzylamine oxidase activity in the conditioned medium from small-scale cultures of HEK293 cells, stably transfected with the GST-SSAO expression vector pMB887, indicated that GST-SSAO was secreted into the culture medium. Further analyses showed that glutathione-Sepharose beads could be used to purify small amounts of the GST-SSAO fusion protein directly from the conditioned medium (data not shown), and that the purified material had benzylamine oxidase activity. Interestingly, the GST-SSAO fusion protein was found to be active also when immobilized on the glutathione-Sepharose beads. The amount of GST-SSAO fusion protein in the conditioned medium was calculated to be ~1 mg/l, by estimation of the amount of protein captured on the beads.

EXAMPLE 5: Preparative purification and site-specific cleavage of the GST-SSAO fusion protein

An overview of the affinity purification based procedure is shown in Fig. 2. The results of the purification are summarized in Table 1. One selected clone was expanded and grown in Cell Factories to generate larger amounts of GST-SSAO for purification. The harvested conditioned medium were concentrated and filtrated to reduce the time for loading on the glutathione-Sepharose column. Glutathione-affinity chromatography was then applied to purify the GST-SSAO fusion protein from the concentrated and filtered conditioned medium. Proteins captured on the column were eluted with 20 mM GSH and analyzed by SDS-PAGE under reducing conditions. This showed that the GST-SSAO fusion protein had high purity and that it could be isolated from large amounts of other proteins in the culture medium in a single step. The GST-SSAO fusion protein migrated in level with the 116 kDa protein in the molecular weight marker. In total 8.8 mg of protein was recovered from the glutathione-Sepharose column. The specific activity of the GST-SSAO fusion protein was determined to 343 nmol · min^{-1} · mg^{-1}. Interestingly, the specific activity was almost doubled (634 nmol · min^{-1} · mg^{-1}) by the buffer exchange step which removed the reducing agent GSH.

The glutathione-affinity purified GST-SSAO was cleaved with the GST-3C protease fusion protein (46 kDa) to remove the GST fusion partner from SSAO. Analytical experiments suggested that cleavage was slow, but precise, with no observable side-products. Moreover, complete cleavage could be obtained after ~48 hours incubation. The cleavage mixture was passed over the glutathione-Sepharose column to capture the removed GST fusion partner and the GST-3C protease. Flow-through material was collected and analyzed by SDS-PAGE under reducing conditions which showed only the expected SSAO product with a molecular weight of ~97 kDa. Captured material was also analyzed, which showed only the GST fusion partner (~28 kDa) and the GST-3C protease. This indicated that a complete cleavage had occurred and all GST containing proteins had been captured on the glutathione-Sepharose column. It also indicated that

all SSAO protein had passed through the column since no SSAO protein was seen in the eluted material.

The specific activity of the purified SSAO protein was determined to 522 nmol · min^{-1} · mg^{-1} which was less than the specific activity determined before cleavage. Since DTT had been used to ensure 3C protease activity during cleavage of the GST-SSAO fusion protein, we made an SDS-PAGE analysis (non-reducing) to see if the cleavage buffer had affected possible disulphide bridges in the SSAO homodimer (Kurkijärvi et al., 1998; Smith et al., 1998; Salminen et al., 1998). Only presumed SSAO monomers (~97 kDa) could be seen (data not shown). However, the SSAO protein was transformed to ~170 kDa in size (analyzed by SDS-PAGE) during storage at 5°C, indicating that one or several disulphides were formed. The cleavage buffer was removed by diafiltration and SDS-PAGE analysis showed that the SSAO protein was still apparently dimeric with a molecular weight of ~170 kDa. In total 3.6 mg of recombinant SSAO was obtained from 9.4 liters of conditioned medium having a specific activity of 809 nmol · min^{-1} · mg^{-1}. The overall yield in the process was 22 % based on determined benzylamine oxidase activity.

Interestingly, the GST fusion partner did not significantly affect the benzylamine oxidase activity of the SSAO protein. The specific activity of the purified GST-SSAO fusion protein after the buffer exchange step, was determined to 634 nmol · min^{-1} · mg^{-1}. After removal of the GST fusion partner, the specific activity of SSAO was determined to 809 nmol · min^{-1} · mg^{-1}. However, the molecular mass of the GST fusion partner is ~25 % of the GST-SSAO fusion protein and the increase in specific activity after removal of GST was in the same range. This opens up possibilities to use the fusion protein for enzyme characterization. Furthermore, an affinity fusion partner such as GST can be used to bind or immobilize a recombinant protein in a directed manner on solid supports to study e.g. protein-protein interactions and enzyme characteristics (Nilsson et al., 1997). The GST-SSAO fusion protein was indeed active when it was bound to glutathione-Sepharose beads.

EXAMPLE 6: Initial characterization of purified SSAO proteins

A gel filtration experiment was performed to analyze the size of the SSAO protein under non-denaturing conditions. A sample from the SSAO protein material that migrated as a dimeric protein when investigated by SDS-PAGE under non-reducing conditions was loaded on a calibrated analytical Superdex 200 column. The SSAO protein eluted at 1.29 ml, which was slightly faster than catalase (232 kDa), which eluted at 1.35 ml.

N-terminal amino acid sequencing of the purified SSAO protein showed that the GST-3C protease had specifically cleaved the 3C protease substrate sequence EALFQG in the GST-SSAO fusion protein (Fig. 1). Twenty-nine amino acids were determined and corresponded exactly to residues number 29-58 in the predicted SSAO amino acid sequence (SEQ ID NO: 2). N-terminal sequencing was also performed on the GST-SSAO fusion protein, which showed that the signal peptide had been processed as anticipated.

Finally, the purified SSAO protein was found to be sensitive to inhibition by semicarbazide as expected. In the presence of 0.1 mM semicarbazide more than 95 % of the benzylamine oxidase activity was inhibited.

TABLE I

Purification of recombinant human SSAO

Purification step (sample)	Total volume (ml)	Total protein (mg)	Total SSAO activity[a] (nmol min^{-1})	Specific activity (nmol min^{-1} mg^{-1})	Yield (%)
Conditioned medium	9400	9024	9243	1.0	—
Concentrated medium	600	9060	13200	1.5	100
Filtrate	850	9065	11900	1.3	90
GSH-affinity step-1 (eluate)	6.8	8.8	3029	343	23
Buffer exchange	15	8.9	5624[b]	634[b]	43
GSH-affinity step-2 (flow-through)[c]	36	5.5	2859	522	22
Diafiltrate[d]	4.2	3.6	2919	809	22

[a] Activity was measured as nmol of H_2O_2 produced per minute using 0.5 mM benzylamine as substrate. SSAO activity was confirmed with 0.1 mM semicarbazide as inhibitor.
[b] Measured before addition of 5 mM DTT and GST-3C protease.
[c] This step was performed after site-specific cleavage of the GST-SSAO fusion protein to capture removed GST fusion partner and GST-3C protease.
[d] Buffer exchange and sample concentration by ultra-filtration.

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CLAIMS

1. A recombinant construct comprising a nucleotide sequence encoding a fusion protein comprising
(i) a soluble form of human SSAO;
(ii) a secretable fusion partner enabling dimerization of SSAO;
(iii) a signal peptide allowing for secretion of a polypeptide from a host cell into the culture medium; and
(iv) a protease cleavage site located between the human SSAO variant and the fusion partner.

2. The method according to claim 1 wherein the said soluble form of human SSAO is lacking the membrane spanning portion essentially shown as amino acids 6 to 26 in SEQ ID NO: 2.

3. The method according to claim 2 wherein the said soluble form of human SSAO essentially consists of amino acids 29 to 763 of SEQ ID NO: 2.

4. The recombinant construct according to any one of claims 1 to 3, encoding a fusion protein wherein the said fusion partner is fused to the N-terminal portion of the human SSAO polypeptide.

5. The recombinant construct according to any one of claims 1 to 4 wherein the said fusion partner is glutathione S-transferase or a functionally equivalent variant thereof.

6. The recombinant construct according to claim 5 wherein the said fusion partner is a variant of *Schistosoma japonicum* glutathione S-transferase, said variant having at least one of the cysteine residues in positions 85, 138, and 178 replaced by another amino acid residue.

7. The recombinant construct according to any one of claims 1 to 6 wherein the said signal peptide is a mouse IgG1 heavy chain signal peptide.

8. The recombinant construct according to any one of claims 1 to 7 wherein the said protease cleavage site is a 3C protease cleavage site.

9. The recombinant construct according to claim 8 wherein the said 3C protease cleavage site comprises the amino acid sequence EALFQG.

10. The recombinant construct according to any one of claims 1 to 9 comprising nucleotide sequence encoding essentially the amino acid sequence shown in Figure 1.

11. An expression vector comprising the recombinant construct according to any one of claims 1 to 10.

12. The expression vector according to claim 11, which is the expression vector termed pMB887, shown in Figure 3.

13. A method for the purification of a recombinant human SSAO polypeptide comprising the steps of:
(i) transfecting cells with an expression vector according to claim 11;
(ii) culturing the said cells under conditions allowing for the fusion protein expressed by the vector to be secreted into the culture medium;
(iii) binding the obtained fusion protein to a medium comprising a ligand having affinity for the fusion partner;
(iv) separating the said fusion partner and the SSAO polypeptide; and
(v) recovering the purified human SSAO polypeptide.

14. The method according to claim 13 wherein the said ligand having affinity for the fusion partner is glutathione, or a derivative thereof.

15. The method according to claim 13 or 14 wherein the said fusion partner is separated from human SSAO in step (iv) by protease cleavage.

16. The method according to claim 15 wherein the said protease is a picornavirus 3C-protease.

17. The method according to claim 16 wherein the said protease is rhinovirus 3C-protease.

18. The method according to claim 16 or 17 wherein the said protease is fused to a fusion partner, thereby obtaining a "fusion protease".

19. The method according to claim 18 wherein the said fusion protease is separated from the SSAO polypeptide by a process comprising binding the said fusion protease to a medium comprising a ligand having affinity for the said fusion protease.

20. A method for the preparation of an immobilized recombinant human SSAO polypeptide comprising the steps of:
(i) transfecting cells with an expression vector according to claim 11;
(ii) culturing the said cells under conditions allowing for the fusion protein expressed by the vector to be secreted into the culture medium;
(iii) binding the obtained fusion protein to a medium comprising a ligand having affinity for the fusion partner.

21. An immobilized recombinant human SSAO polypeptide obtainable by the method as defined in claim 20.

22. An immobilized or encapsulated recombinant human SSAO polypeptide obtainable by the method as defined in claim 13.

ABSTRACT

The present invention relates to a recombinant construct comprising a nucleotide sequence encoding a fusion protein comprising a soluble form of human SSAO (Semicarbazide-Sensitive Amine Oxidase), a secretable fusion partner, a signal peptide; and a protease cleavage site. The said construct is useful in methods for purification of a soluble form of human SSAO.

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Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu Met
115 120 125

Leu Lys Met Phe Glu Asp Arg Leu Ser His Lys Thr Tyr Leu Asn Gly
130 135 140

Asp His Val Thr His Pro Asp Phe Met Leu Tyr Asp Ala Leu Asp Val
145 150 155 160

Val Leu Tyr Met Asp Pro Met Cys Leu Asp Ala Phe Pro Lys Leu Val
165 170 175

Ser Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr Leu
180 185 190

Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala Thr
195 200 205

Phe Gly Gly Gly Asp His Pro Pro Lys
210 215

Fig. 1

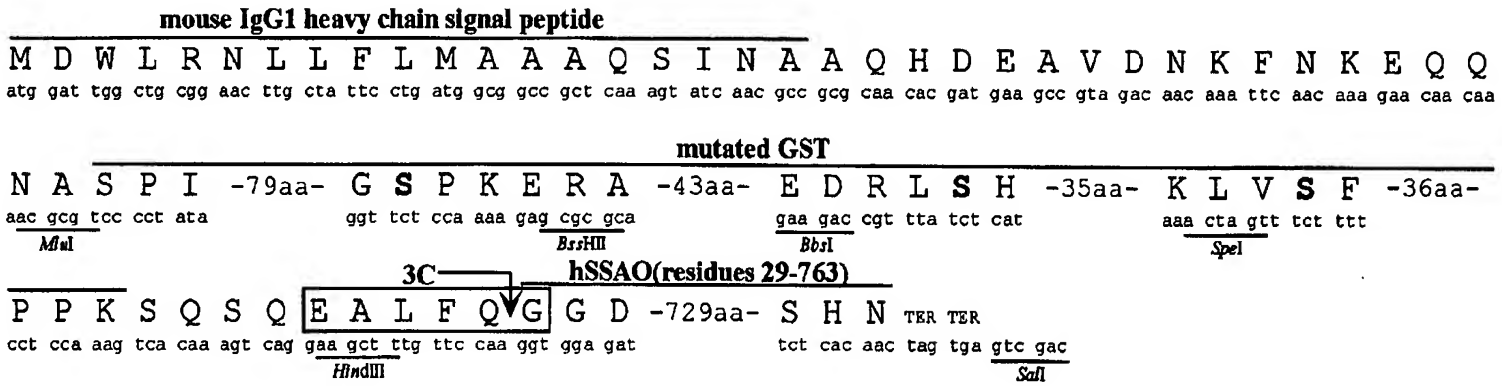
mouse IgG1 heavy chain signal peptide
M D W L R N L L F L M A A A Q S I N A A Q H D E A V D N K F N K E Q Q
atg gat tgg ctg cgg aac ttg cta ttc ctg atg gcg gcc gct caa agt atc aac gcc gcg caa cac gat gaa gcc gta gac aac aaa ttc aac aaa gaa caa caa
mutated GST
N A S P I -79aa- G S P K E R A -43aa- E D R L S H -35aa- K L V S F -36aa-
aac gcg tcc cct ata ggt tct cca aaa gag cgc gca gaa gac cgt tta tct cat aaa cta gtt tct ttt
MfuI
BssHII
BbsI
SpeI
3C
hSSAO(residues 29-763)
P P K S Q S Q E A L F Q G G D -729aa- S H N TER TER
cct cca aag tca caa agt cag gaa gct ttg ttc caa ggt gga gat tct cac aac tag tga gtc gac
HindIII
SalI

Fig. 2

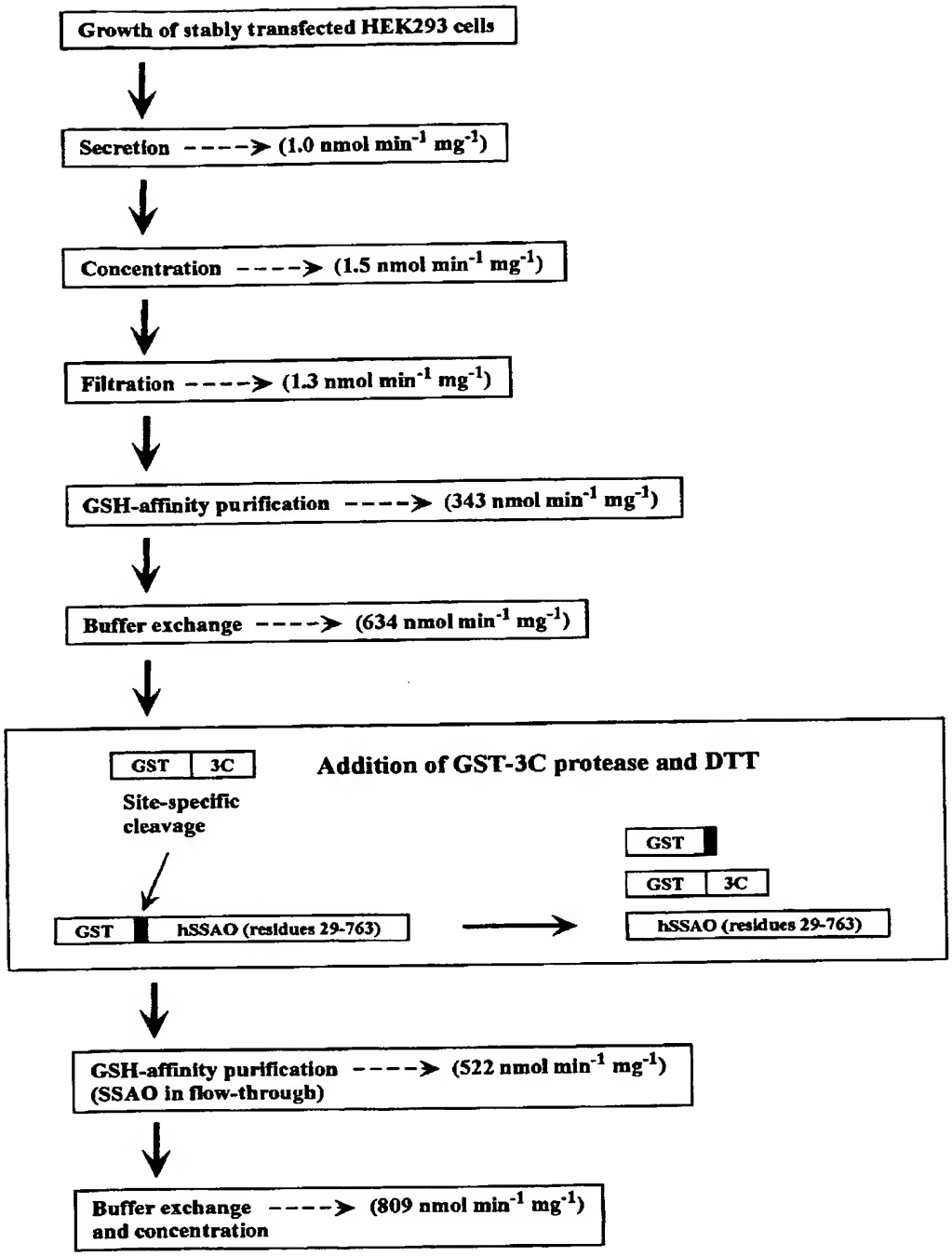

Fig. 3